Suitability of VACUETTE® FC Mix blood collection tubes for gestational diabetes testing

**Background:**
Greiner-Bio-One, Austria has been selling plastic evacuated tubes (VACUETTE®) for venous blood collection since 1986.

VACUETTE® FC Mix blood collection tubes contain an additive mix of Na₂EDTA, sodium fluoride, citric acid and sodium citrate. This mixture inhibits glycolysis and prevents coagulation. [1][2]

The VACUETTE® FC Mix blood collection tubes are used to stabilize the in-vivo glucose level in whole blood or plasma for up to 48 hours at room temperature and enable prolonged sample processing time including both storage and/or transportation. [3]

Due to the incomplete inhibition of the glycolysis by sodium fluoride, new German guidelines recommend the usage of tubes containing the additive mixture of fluoride, citric acid and citrate. Furthermore, critical values in order to assess the glucose metabolism and diagnose diabetes mellitus and gestational diabetes are defined by those guidelines. [4][5]

Gestational diabetes
- Fasting plasma glucose > 92 mg/dL (> 5.1 mmol/L)
- OGTT-1h-value > 180 mg/dL (> 10.0 mmol/L)
- OGTT-2h-value > 153 mg/dL (> 8.5 mmol/L)

**Study Objective:**
The study has been carried out to demonstrate that VACUETTE® FC Mix blood collection tubes are suitable for gestational diabetes diagnosis of pregnant women by stabilization of glucose concentration for up to 48h after blood collection in comparison to Terumo VENOSAFE™ FC Mixture.

**Study design and procedure:**
Venous whole blood was collected from 43 pregnant donors who were healthy (n=19) or diagnosed with gestational diabetes (n=24) by using a VACUETTE® SAFETY Blood Collection Set (Item #450085) into the following tubes:

**Sample A:** VENOSAFE™ FC Mixture® (Prod. No. VF-052SCF)
**Sample B:** VACUETTE® FC Mix (Prod. No. 454511)

All samples were centrifuged at 1800g for 10 min at 20°C (centrifuge: Eppendorf 5810R). In order to test the robustness of whole blood, two tubes of each sample were taken. One of each sample was centrifuged immediately after blood collection. The second tube was split into four portions and centrifuged under three different time periods:
1) 1h after blood collection and
2) 24h after blood collection
3) 48h after blood collection

**Time scheme:**

**Blood collection**
- **Tube 1:**
  - A1/C1 0h centrifugation and initial value measurement
  - A1/C1 1h 1h measurement
  - A1/C1 12h measurement
  - A1/C1 24h 24h measurement
  - A1/C1 48h 48h measurement

- **Tube 2:**
  - A2/C2 0h centrifugation and initial value measurement
  - A2/C2 1h 1h measurement
  - A2/C2 24h 24h measurement
  - A2/C2 48h centrifugation and 48h measurement
The sample tubes which were centrifuged initially after blood collection were analyzed for glucose at the initial time point, after 1h, 12h, 24h and 48h after blood collection on the COBAS 8000 analyzer (Roche diagnostics, Mannheim). The sample tubes which were stored up to 48h after blood collection at room temperature were centrifuged and analyzed according to the time scheme given above. From each sample, the hemolytic index was determined on the COBAS 8000 as well.

Between measurements, all samples were stored in an upright position at room temperature.

One value is missing (A3, Donor 21) due to an analyzer error.

**Results:**

1. **Centrifugation directly after blood collection**

The mean values of the glucose concentration for the samples which were centrifuged directly after blood collection are shown in Table 1.

<table>
<thead>
<tr>
<th>Sample</th>
<th>Valid N</th>
<th>Mean</th>
<th>±SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>A1 0h: Terumo VENOSAFE™ FC Mixture initial</td>
<td>43</td>
<td>6.81</td>
<td>1.76</td>
</tr>
<tr>
<td>A1 1h: Terumo VENOSAFE™ FC Mixture</td>
<td>43</td>
<td>6.78</td>
<td>1.76</td>
</tr>
<tr>
<td>A1 12h: Terumo VENOSAFE™ FC Mixture</td>
<td>43</td>
<td>6.77</td>
<td>1.81</td>
</tr>
<tr>
<td>A1 24h: Terumo VENOSAFE™ FC Mixture</td>
<td>43</td>
<td>6.92</td>
<td>1.79</td>
</tr>
<tr>
<td>A1 48h: Terumo VENOSAFE™ FC Mixture</td>
<td>43</td>
<td>6.93</td>
<td>1.77</td>
</tr>
<tr>
<td>C1 0h: Greiner VACUETTE® FC Mix initial</td>
<td>43</td>
<td>6.76</td>
<td>1.74</td>
</tr>
<tr>
<td>C1 1h: Greiner VACUETTE® FC Mix</td>
<td>43</td>
<td>6.73</td>
<td>1.72</td>
</tr>
<tr>
<td>C1 12h: Greiner VACUETTE® FC Mix</td>
<td>43</td>
<td>6.77</td>
<td>1.73</td>
</tr>
<tr>
<td>C1 24h: Greiner VACUETTE® FC Mix</td>
<td>43</td>
<td>6.82</td>
<td>1.74</td>
</tr>
<tr>
<td>C1 48h: Greiner VACUETTE® FC Mix</td>
<td>43</td>
<td>6.83</td>
<td>1.74</td>
</tr>
</tbody>
</table>

Comparison analysis was performed at all time points of determination. Statistics was performed with the t-test (α = 0.05) using StatSoft Software, Version 12.

Clinical evaluation was based on the allowed recommendation by the German Medical Association (RILIBÄK). [8].

![Graph](image)

**Figure 1:** Initial, 1h, 24h and 48h values
Tube 1: specimen centrifuged immediately after blood collection, replicate measurements after 1h, 12h, 24h and 48h.
Figure 1 illustrates the initial, 1h, 24h and 48h values for both samples centrifuged directly after blood collection. The stabilization of the glucose concentration has been demonstrated in both samples, no clinically or statistically significant deviations have been found comparing sample A1 to C1 at each point of time (highest single deviations: 5.50% initial, 4.98% after 1h, 6.08% after 12h, 6.07% after 24h and 6.40 after 48h). The highest deviation when comparing the time points has been found for sample A1 after 48h (A1/0h to A1/48h: -5.37% and C1/0h to C1/48h: 6.56%).

On the basis of the healthy or with gestational diabetes pregnant women tested, the equivalence in the performance of the samples that were centrifuged directly after blood collection has been shown.

The hemolytic index in the tubes, that were immediately centrifuged was very low (Hemolytic index <50 mg/dl). Storage of whole blood samples increases the likelihood of hemolysis as found for some samples centrifuged later (Hemolytic index <250 mg/dl), however without any impact on the determination of glucose concentration on the COBAS 8000 analyzer.

2. Storage of whole blood samples

Table 2 summarizes the results of samples stored as whole blood without immediate centrifugation.

<table>
<thead>
<tr>
<th>Sample</th>
<th>Valid N</th>
<th>Mean</th>
<th>±SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>A2 0h: Terumo VENOSAFE™ FC Mixture initial</td>
<td>43</td>
<td>6.78</td>
<td>1.76</td>
</tr>
<tr>
<td>A3 1h: Terumo VENOSAFE™ FC Mixture</td>
<td>42</td>
<td>6.80</td>
<td>1.81</td>
</tr>
<tr>
<td>A4 24h: Terumo VENOSAFE™ FC Mixture</td>
<td>43</td>
<td>6.77</td>
<td>1.82</td>
</tr>
<tr>
<td>A5 48h: Terumo VENOSAFE™ FC Mixture</td>
<td>43</td>
<td>6.77</td>
<td>1.80</td>
</tr>
<tr>
<td>C2 0h: Greiner VACUETTE® FC Mix initial</td>
<td>43</td>
<td>6.76</td>
<td>1.74</td>
</tr>
<tr>
<td>C3 1h: Greiner VACUETTE® FC Mix</td>
<td>43</td>
<td>6.74</td>
<td>1.78</td>
</tr>
<tr>
<td>C4 24h: Greiner VACUETTE® FC Mix</td>
<td>43</td>
<td>6.69</td>
<td>1.79</td>
</tr>
<tr>
<td>C5 48h: Greiner VACUETTE® FC Mix</td>
<td>43</td>
<td>6.68</td>
<td>1.78</td>
</tr>
</tbody>
</table>

Figure 2: Storage as whole blood
Tube 2: specimen centrifuged after 1h (A3/C3), 24h (A4/C4) or 48h (A5/C5) after blood collection.
The stability of the glucose concentration in sample C2 (48h in whole blood) has been demonstrated as shown in Figure 2. The comparison between the initial values of sample C1 and the 48h values of sample C2 did not result in statistically or clinically significant differences due to sodium fluoride acting as an enolase inhibitor being responsible for the long term stabilization of the glucose concentration.

The stability of the glucose concentration in samples A3, A4, A5 as well as C3, C4, C5 (2h, 24h and 48h stored as whole blood) has been demonstrated by Figure 2. The highest deviations have been found for the comparison of A2/0h to A5/48h with 8.21% and C2/0h to C5/48h with 10.09%.

These results lead to the conclusion that storage for whole blood samples also illustrates the equivalence in the performance of both blood collection tubes used for comparison testing.

**Conclusion:**

Using blood collection tubes containing the additive combination of citrate, EDTA and fluoride, acting as an enolase inhibitor being responsible for the long term stabilization of the glucose concentration, the stabilization of glucose concentration in the VACUETTE® FC Mix blood collection tubes in comparison to VENOSAFE™ FC Mixture (Terumo) was demonstrated for up to 48h at room temperature for pregnant donors who were healthy or diagnosed with gestational diabetes. No clinically significant differences were observed between VENOSAFE™ FC Mixture and VACUETTE® FC Mix tube at any time point when stored in an upright position. Furthermore, stability of glucose concentration was shown regardless of delayed centrifugation time up to 48h after the blood collection. These results indicate the suitability of VACUETTE® FC Mix blood collection tubes for reliable determination of glucose concentration for pregnant women if prolonged processing times including transport and/or storage times occur. Further research with a larger group could be done to further sustain this conclusion. On the basis of utilization of VACUETTE® FC Mix Blood collection Tube, an impact of hemolytic index up to 250 mg/dl on the glucose concentration could not be proven.

**References:**


