

# First experiments on the possible use of diphenhydramine as a model substance for the evaluation of oral fluid sample collection

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# Introduction

Oral fluid (OF) is gaining increasing interest in drugs of abuse and compliance testing as a less intrusive matrix compared to serum or urine. However, little is known about the influence of the collection process on analyte recovery depending on different collection devices. In addition contamination of OF with the corresponding drugs shortly after drug ingestion may lead to interpretation problems. Furthermore unintended oral contamination (eg. "kissing") could be cited (falsely or correctly) by patients / clients to explain their positive drug testing results. A proper model substance to investigate possible ways of oral contamination and the OF sampling process with its influencing factors is therefore needed. Pharmacological and physicochemical properties (pKa: 9.0 plasma protein binding: 80%, oral bioavailability: 50% - 70%, t<sub>1/2</sub>: 4h), relative safety and availability (non-prescription drug) makes Diphenhydramine (DPH) a candidate substance for evaluation. This approach is complemented by the ease of capillary whole blood sampling.

# Methods

**Patients:** All volunteers were between 23 and 47 years of age, some participated in more than one

Sample collection: OF samples were collected using the liquid based Greiner Bio-One (GBO, Kremsmünster, Austria) SCS pH 4.2 device or the Quantisal (Immunalysis, Pomona, USA) device according to the manufacturer. Capillary whole blood (CB) was collected with the Minivette POCT (Sarstedt, Nümbrecht, Germany) according to the manufacturer. Sample collection was performed by the volunteers themselfes. All collection devices used in the study are approved for self application by the manufacturer. Amylase and oral fluid concentration in GBO samples were measured with an Olympus

Sample preparation: 20µl CB or OF/SES fortified with 20 µL internal standard (100 ng/mL DPH-d<sub>3</sub> in MeOH), was protein precipitated with 60 μL MeOH/ACN (50:50, v/v). After centrifugation 20 μL of the supernatant was diluted with 130 μL MeOH/H<sub>2</sub>O (60:40, v/v) + 0.1% FA. 5 μL were injected into the UPLC system. CB and OF/SES matrix calibration was performed from 1 to 1000 ng/mL (LoD: OF = 0.34 ng/mL CB = 0.37 ng/mL; LoQ: OF = 0.42 ng/mL, CB = 0.48 ng/mL).

**UPLC-MS/MS:** Gradient separation was conducted on a Waters Acquity UPLC connected to a Xevo-TQ-S with a BEH Phenyl column (1.7 μm, 2.1x100 mm), kept at 40 °C within 2.5 min. MoPh A was 0.1% FA in H<sub>2</sub>O and MoPh B was 0.1% FA in MeOH. The instrument operated in ESI positive mode and two transitions for each analyte were recorded.

# Conclusion

The detection times of DPH in OF and CB are comparable.

-- The DPH OF/CB ratio of the first two hours after ingestion (mean: 5.96, CV: 113%, range: 1.13 - 23.8, n= 12) was significantly higher than during elimination (4 - 58 h; mean: 1.62, range: 0.27 - 4.09; CV: 60%, n = 47) indicating OF contamination from the uncoated

After induced contamination of OF with DPH, high drug concentrations could be detected (49947 - 296553 ng/mL OF/SES). Even after rinsing the mouth with 248 ml rinsing solution in twenty steps, within 45 minutes, 0.04 - 0.1% of the initial drug concentration was detectable in the rinsing solution (20.5 - 285 ng/mL OF/SES; see Fig. 5, 6). Both collection devices showed reproducible recovery after consecutive OF sampling in different individuals with the same collection device (Fig. 3). Consecutive sampling with the

two different OF collection devices in the same individuals gave comparable concentrations

(Fig. 4). DPH fulfills the key criteria for a good model substance: long detectability in both matrices. "sufficient" elimination half-life and a correlation between OF and blood concentrations.

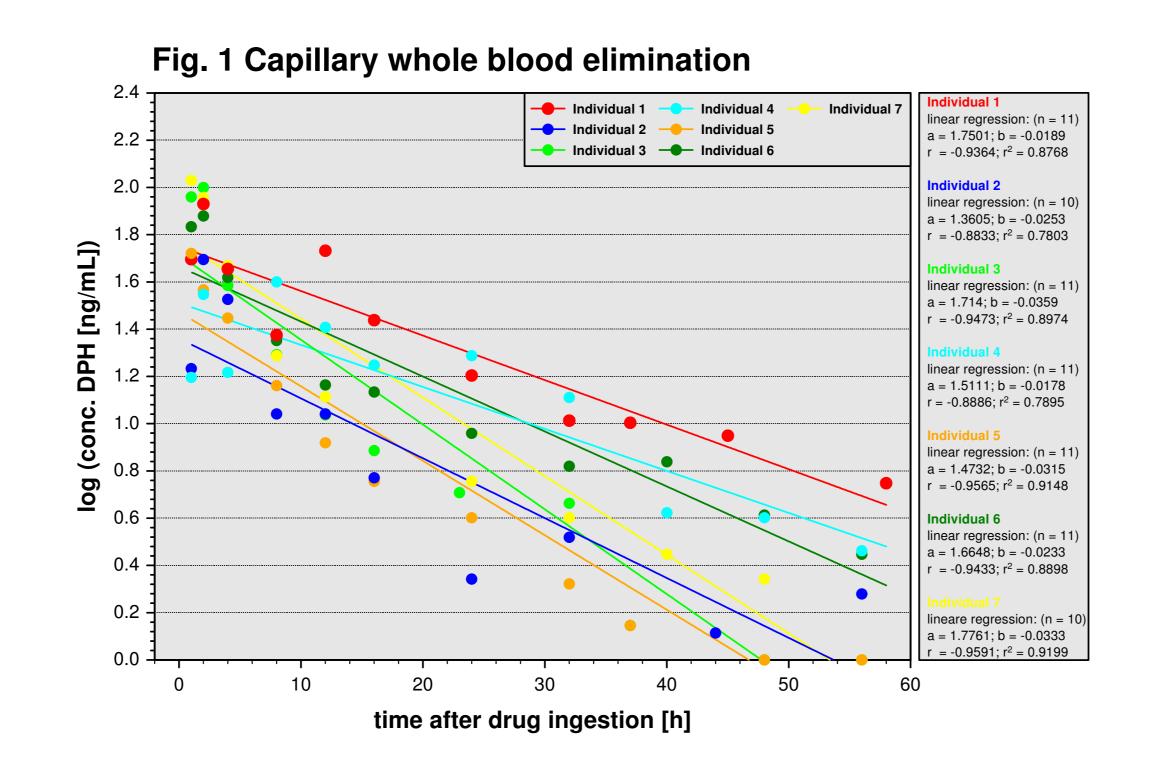
# Results

#### **Experimental design: Elimination of DPH in OF and CB**

Step 1: before drug ingestion, sampling of OF and CB ingestion of one uncoated tablet of DPH-HCI 50 mg Step 2: ("Dorm", Berco, Kleve, Germany) Step 3: susequently, sampling of OF and CB Step 4: rinsing mouth and lips with water for one minute Step 5: subsequently, sampling of CB and OF after 1h, 2h and 4h, sampling of OF and CB Step 6 - 8: sampling of all three matrices every 4 hours until 16h Step 9 - 11: after drug ingestion Step 12 - 16: sampling of all three matrices every 8 hours until 56h after drug ingestion

**Individuals: Collecting devices:** -- CB: Sarstedt Minivette POCT -- male: 4, 23 - 28 years

-- OF: Greiner Bio-One -- female: 3, 23 - 29 years



#### Fig. 2 Oral fluid elimination Individual 4 linear regression: (n = 11)oral contamination a = 1.9124; b = -0.0224r = -0.8898; $r^2 = 0.7917$ a = 1.5502; b = -0.0301r = -0.9378; $r^2 = 0.8794$ linear regression: (n = 11) a = 2.4073; b = -0.039r = -0.9595; $r^2 = 0.9206$ DPH a = 1.9891; b = -0.0385r = -0.9555; $r^2 = 0.9129$ a = 2.3467; b = -0.0523r = -0.9057; $r^2 = 0.8202$ 1.0 a = 2.1596; b = -0.0323r = -0.8987; $r^2 = 0.8076$ 0.5 linear regression: (n = 11) a = 2.0502; b = -0.0411 r = -0.9231; $r^2 = 0.8522$ 0.0 -50

time after drug ingestion [h]

### **Experimental design: Analyte recovery - Greiner vs. Quantisal**

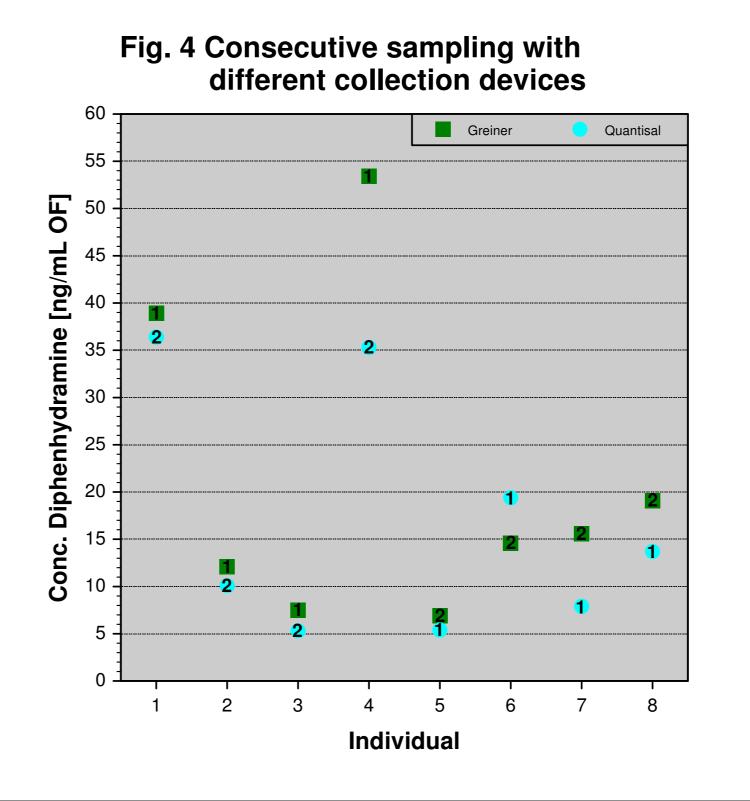
ingestion of one uncoated tablet of DPH-HCI 50mg ("Dorm", Berco, Kleve, Germany) after 12 h collection of 3 OF samples with the Quantisal collection device (Group a) or with the Greiner Bio-One collection device (Group b) within 30 minutes maximum 90 min later one collection of one OF samle with the Step 3: Greiner Bio-One collection device and one with the Quantisal collection device (Group a) respectively the other way around (Group b)

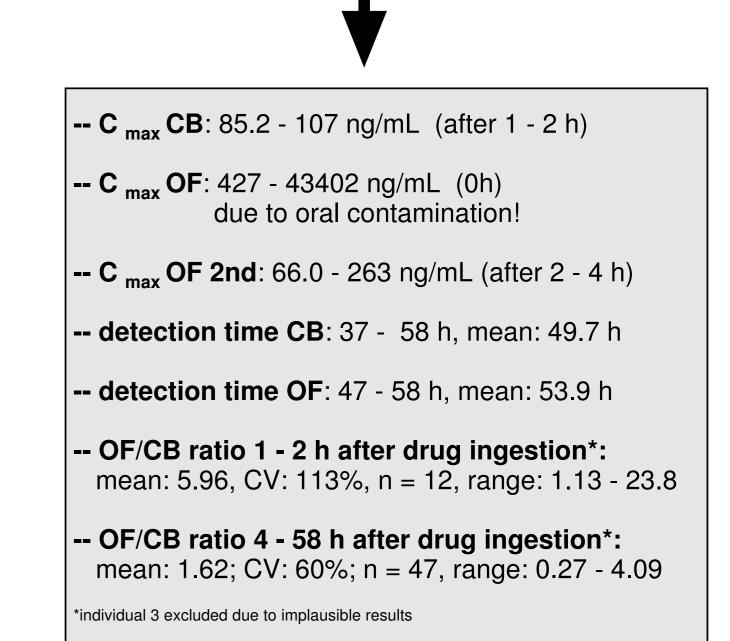
**Collection devices:** Individuals: -- a): Immunalysis Quantisal -- male: 4, 23 - 28 years -- b): Greiner Bio-One -- female: 4, 23 - 29 years

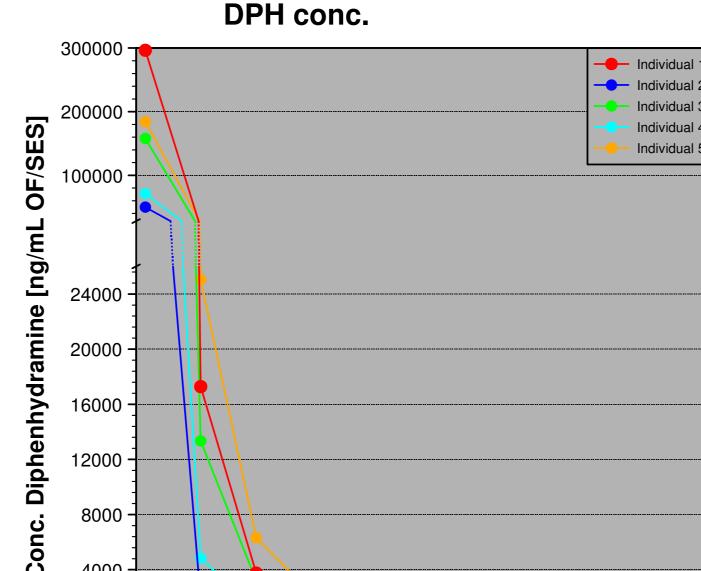
# collection device Quantisal Greiner 1 - 3 2+3 Dip Individua

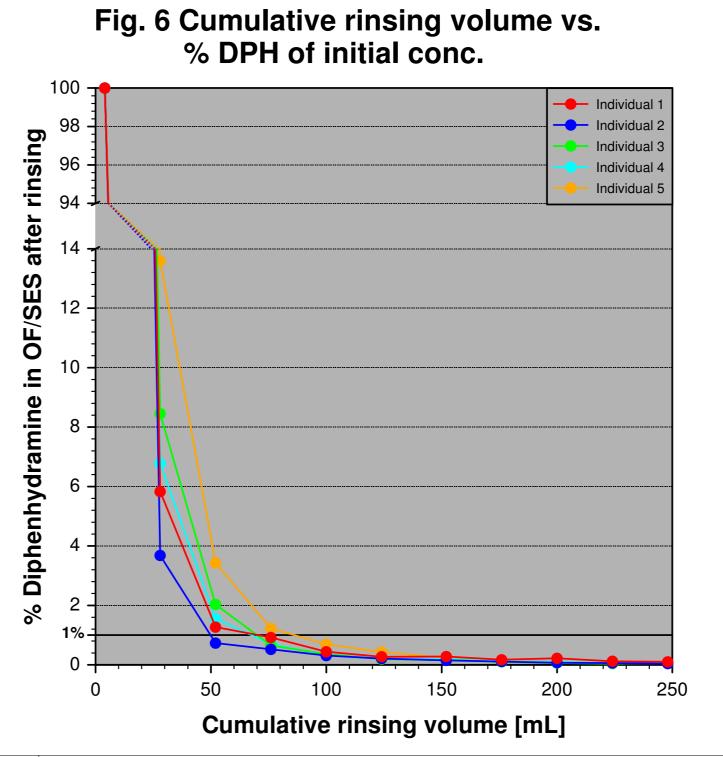
Fig. 5 Cumulative rinsing volume vs.

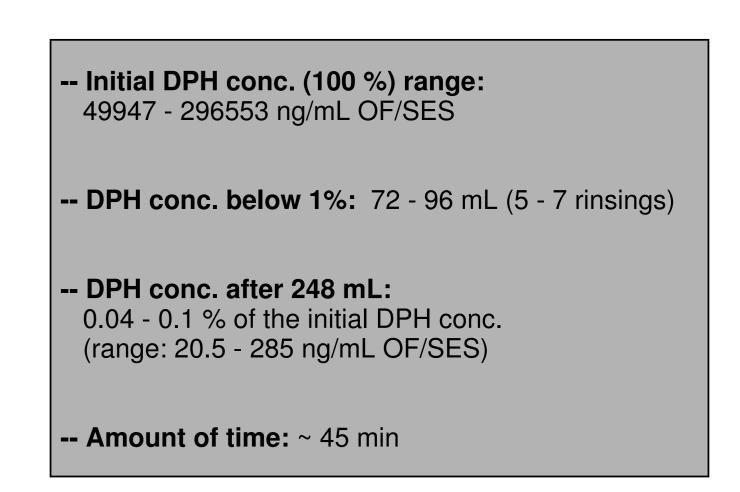
Fig. 3 Consecutive sampling with same











Individuals: Rinsing solutions (RS):

**Experimental design:** 

**Oral contamination** 

sampling of OF before drug ingestion

subsequently sampling of OF with RS a rinsing mouth with RS b for 30 seconds

repeat step 3 and 4 nine times

sampling of OF (RS a)

putting one uncoated tablet of DPH-HCI 50mg

("Dorm", Berco, Kleve, Germany) into the mouth for

Step 1:

Step 2:

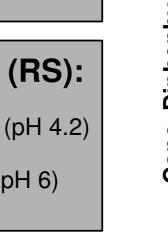
Step 3:

Step 4:

Step 13:

Step 5 - 12:

-- male: 1, 34 years -- RS a: 4 mL SES (GBO) (pH 4.2) -- female: 4, 23 - 47 years -- RS b: 20 mL tap water (pH 6)



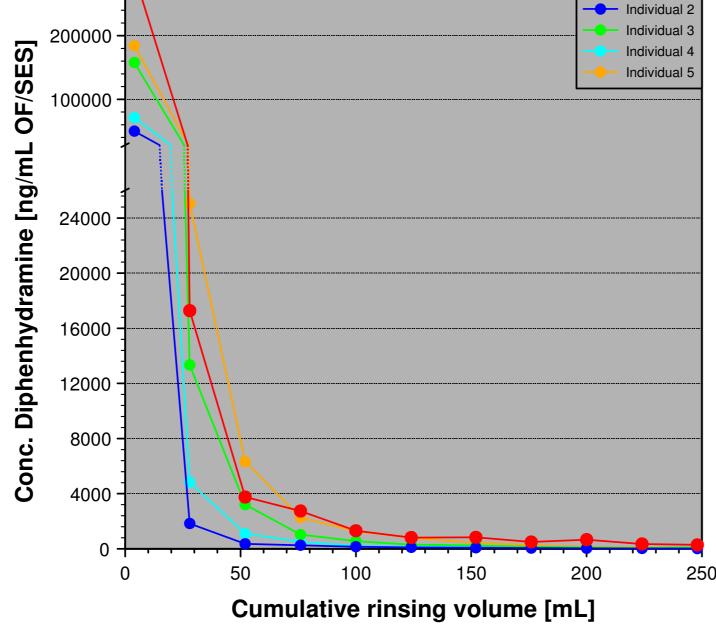
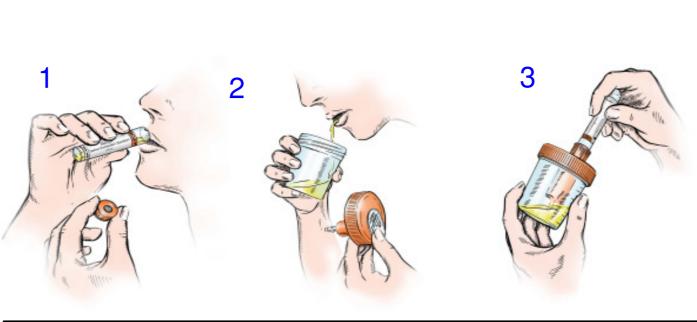


Fig. 7 Greiner Bio-One Saliva Collection System pH 4.2





## Saliva collection

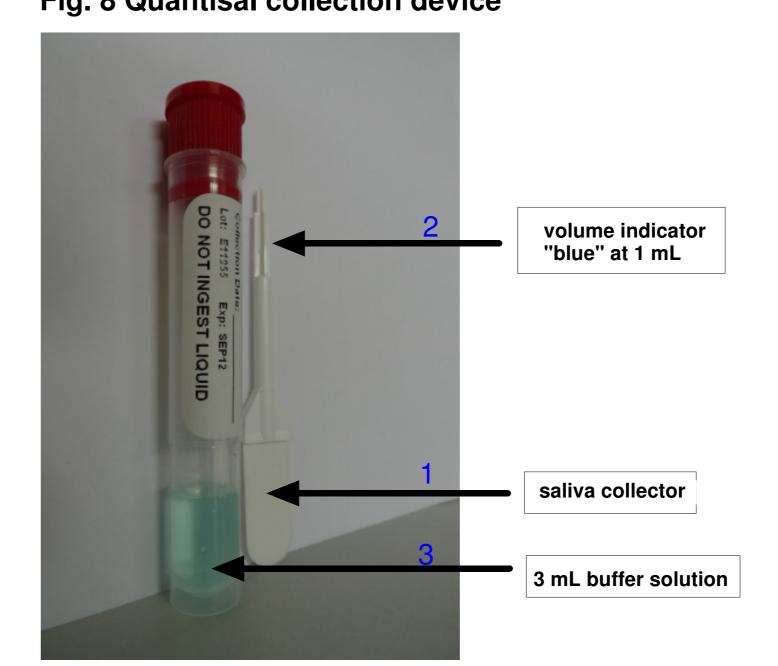
(1) rinse oral cavity with Saliva Extraction Solution (SES) for minimum 2 minutes

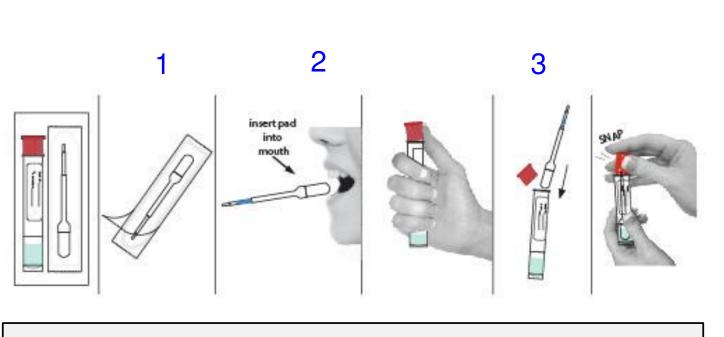
(2) spit OF/SES into beaker (3) transfer OF/SES into evacuated tubes containing bactericides

(4) after centrifugation Amylase and OF concentration are determined

on an Olympus AU680

### Fig. 8 Quantisal collection device





### Saliva collection

(1) put the salica collector under the tongue (2) wait until the volume indicator turns "blue" (3) transfer the saliva collector in the buffer solution and send it the lab

(4) OF concentration is 25%